



Synthesis and antibacterial activities of a novel alkylide: 3-O-(3-aryl-2-propargyl) and 3-O-(3-aryl-2-propenyl)clarithromycin derivatives

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ABSTRACT

A series of novel 3-O-(3-aryl-*E*-2-propenyl)clarithromycin derivatives **8** and 3-O-(3-aryl-2-propargyl)clarithromycin derivatives **11** were designed, synthesized, and evaluated for their in vitro antibacterial activities. Compared with **8c** and **11c** (Ar was 5-pyrimidyl), 3-O-(3-(5'-pyrimidyl)-*Z*-1-propenyl) counterpart **6c** displayed 4- to 64-fold more potent activities against erythromycin-susceptible *Staphylococcus aureus* and *Streptococcus pneumoniae*. Moreover, the activities of **6c**, **8c**, and **11c** against erythromycin-resistant *S. aureus* and *S. pneumoniae* were in general 4-fold higher than those of the reference compound, clarithromycin and azithromycin.

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The second-generation erythromycin, exemplified by clarithromycin (6-O-methylerythromycin, as shown in Fig. 1), was developed to address the acid instability of erythromycin A, which was associated with an intramolecular ketalization of 6-OH with 9-keto.¹ Clarithromycin was successfully synthesized via regioselective methylation at 6-OH of erythromycin.² As a result, clarithromycin exhibits better bioavailability and improved pharmacokinetics.³

Recently, the resistance against erythromycin and clarithromycin is increasingly prevalent via two main genotypes: ribosome methylation or dimethylation resistance encoded by *erm* gene and efflux resistance encoded by *mef* gene.⁴ Meanwhile, the *erm*-mediated resistance exists in two phenotypes: inducible and constitutive resistance.

To combat the resistance, a series of ketolides, derived from erythromycin by substitution of 3-O-cladinose sugar by a 3-keto group, were synthesized and evaluated.⁵ Consequently, ketolides are distinguished by telithromycin (HMR3647),⁶ cethromycin (ABT-773),⁷ and TE-802,⁸ as shown in Fig. 2. Telithromycin and cethromycin were shown to effectively address *erm*- and *mef*-containing resistance, and TE-802 proved to effectively overcome *mef* resistance but possessed weak activities against *erm* resistance. However, the discovery of ketolides disproved the long held belief that 3-O-cladinose was indispensable moiety for the antibacterial activity.

Encouraged by the important breakthrough, a novel acylide characterized by 3-O-acetyl group was prepared after removal of cladinose. Among the candidates, 3-O-(3-pyridyl)acetyl acylide (TEA-0929) possessed better activity against *mef* resistant strains^{9,10} (Fig. 3). In addition to continuous focus on the synthesis of new ketolides and acylides,^{11–13} other macrolides with various substituents at C-3 instead of cladinose have also been designed and investigated over the past decade, such as 3,6-bicyclolide¹⁴ (Fig. 3), 2,3-anhydrolide,¹⁵ 2,3-enol ether,¹⁶ 3-deoxy,¹⁷ 3-O-phenyl ether,¹⁸ 3,6-ketal,¹⁹ and 3,6-ether.²⁰

The structure–activity relationship study indicated that ketolides, acylides, and 3,6-bicyclolides presented good activities against *mef* resistance. Furthermore, an aryl group tethered to macrolides contributed much to overcome *erm* resistance via additional contacts with domain II of bacterial ribosome.

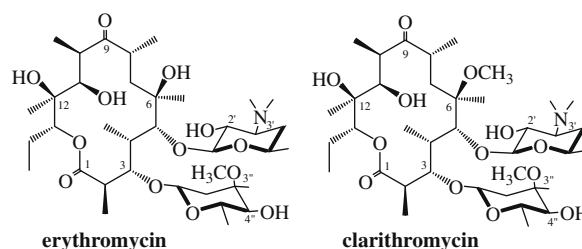


Figure 1. Structure of erythromycin and clarithromycin.

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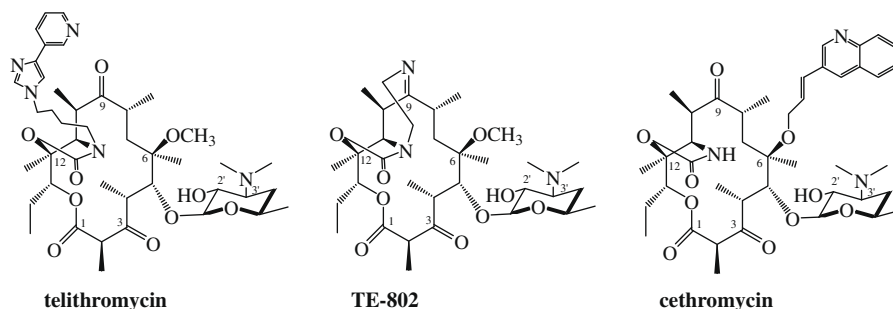


Figure 2. Structure of ketolide.

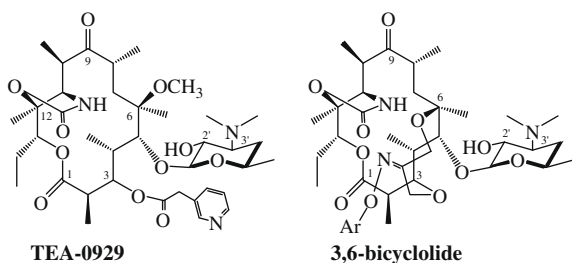
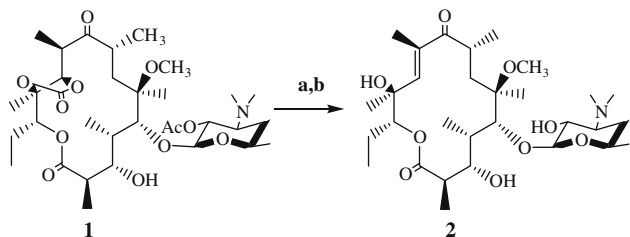


Figure 3. Structure of acylide and 3,6-bicyclolide.

An allyl and a propargyl group have been introduced to 6-OH regioselectively, and the resulting products served as versatile intermediates for a variety of further modification,^{7,21–25} such as the introduction of aryl groups via Heck or Sonogashira reaction. Thus, we presumed that 3-O-allyl clarithromycin and 3-O-propargyl clarithromycin derivatives should be potential lead compounds for further screening. Consequently, we accomplished the regioselective allylation at 3-OH and obtained a series of 3-O-(3-aryl-Z-1-propenyl)clarithromycin derivatives via Heck reaction, which we named alkylides.²⁶ To further study of the structure–activity relationships of alkylides, we herein report the synthesis and the antibacterial activities of 3-O-(3-aryl-E-2-propenyl) and 3-O-(3-aryl-2-propargyl)clarithromycin analogues.

A straightforward approach to 3-O-allyl alkylide failed via allylation of 3-OH-9-keto **1**,²⁷ and instead the major product in the resulting mixture was identified as 3-OH-10,11-anhydro **2**, as illustrated in Scheme 1. In contrast, 3-OH-9-O-(2-chlorobenzyl)oxime **3** served as the key precursor to produce corresponding 3-O-allyl **4** in 95.2% yield,²⁶ as presented in Scheme 2.

Interestingly, the Heck reaction of 3-O-allyl **4**, in the presence of palladium(II) acetate and tri(*o*-tolyl) phosphine as catalysts, easily resulted in the allylic double bond isomerization from 2-position to 1-position as well as the configuration isomerization from *E* to *Z* (**5** and **6**),²⁶ presumably due to steric reasons in β -hydrogen elimination (Scheme 2). However, further study in our lab disclosed that the catalyst could efficiently give the desirable sidechain without



Scheme 1. Reagents and conditions: (a) allyl bromide, KOtBu, THF/DMSO, 0 °C, 50% (TLC); (b) MeOH, then purified by column chromatography, 11.7% over two steps.

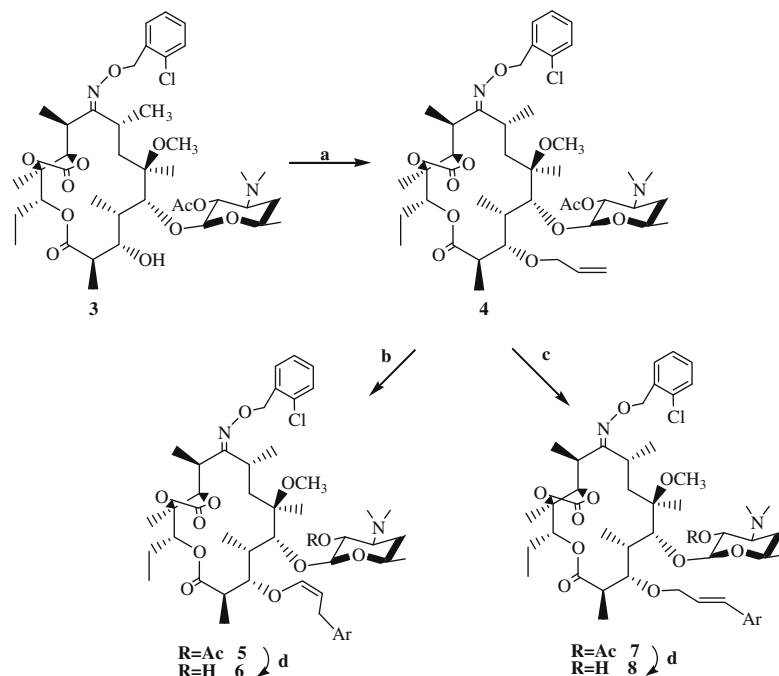
Heck isomerization under the precondition of 2,3-dehydro-3-O-allyl analogues, and the results in detail will be published elsewhere. Thus, the Heck reaction of 3-O-allyl clarithromycin derivatives may warrant further study.

To prevent the unexpected Heck isomerization of **4**, a new kind of palladacycle catalyst, *trans*-di(μ -acetato)bis[O-(di-*o*-tolylphosphino)benzyl]dipalladium(II)²⁸ was utilized in our synthesis of the alkylides (**7** and **8**), as presented in Scheme 2. The catalyst was extensively used in the synthesis of 3-O-aryl-E-2-propenyl leucomycin analogues, a 16-membered macrolide.²⁹ As expected, the application of the palladacycle catalyst successfully furnished the desirable 3-O-[3-(3'-pyridyl)-E-2-propenyl] (**8b**) and 3-O-[3-(5'-pyrimidyl)-E-2-propenyl] sidechain (**8c**). Nonetheless, it was noted that the palladacycle catalyst could not afford 3-O-[3-(4'-isoquinolyl)-E-2-propenyl] sidechain (**8e**) efficiently, and the isomerization product (**6e**) still proved to be the majority, as was attributed to the bulky hindrance of 4-isoquinolyl group.

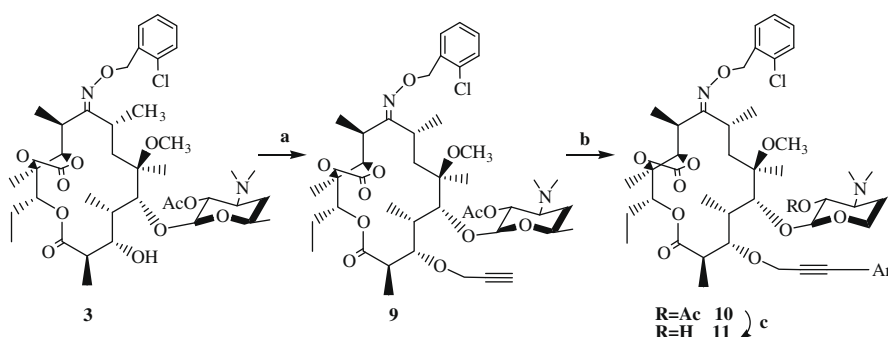
As for the introduction of a propargyl group at 3-OH, it was first reported by Omura group that treatment of 3-OH-6,9-hemiketal derivative, accessible from the acid degradation product of erythromycin, with propargyl bromide and NaH gave 3-O-propargyl-6,9-hemiketal counterpart in 72% yield.³⁰ However, no reaction and activities were reported concerning the Sonogashira coupling products. To further explore the effect of the sidechain's configuration on the activities, the key intermediate 3-O-propargyl alkylide **9** was synthesized, in the presence of propargyl bromide and KOtBu, from the precursor 3-OH-9-O-(2-chlorobenzyl)oxime **3** in 89.5% yield. Next, treatment of **9** with a variety of aryl halides under the Sonogashira coupling conditions efficiently led to the expected coupling products **10**. The aromatic rings included 4-nitrophenyl (**a**), 3-pyridyl (**b**), 5-pyrimidyl (**c**), 3-quinolyl (**d**), 4-isoquinolyl (**e**), 5-indolyl (**f**) group. Finally, hydrolysis of the 2'-O-Ac by heating in methanol provided the desired **11** (Scheme 3).

The antibacterial activities of 3-O-(3-aryl-Z-1-propenyl) alkylide **6**, 3-O-(3-aryl-E-2-propenyl) alkylide **8** and 3-O-(3-aryl-2-propargyl) alkylide **11** were assessed against a panel of erythromycin-susceptible and erythromycin-resistant bacteria. The selected strains are *Staphylococcus aureus* ATCC 29213 (erythromycin-susceptible strain), *S. aureus* PU-32 (erythromycin-resistant strain bearing inducible *erm* (A) gene), *S. aureus* PU-19 (erythromycin-resistant strain bearing constitutive *erm* (A) gene), *Streptococcus pneumoniae* ATCC 49619 (erythromycin-susceptible strain), *S. pneumoniae* PU-11 (erythromycin-resistant strain bearing both *erm* and *mef* genes), and *S. pneumoniae* PU-13 (erythromycin-resistant strain bearing both *erm* and *mef* genes). Data are listed in Table 1 as the minimal inhibitory concentration (MIC), which is determined by the broth microdilution method as recommended by Clinical and Laboratory Standards Institute (CLSI).³¹

Actually, the activities of the alkylides **6**, **8**, **11** were comparable to those of the corresponding 9-O-(2-chlorobenzyl)oxime ketolides³² and some triazole derivatives started from 3-O-propargyl-6,9-hemiketal.³⁰ The alkylides exhibited improved activities



Scheme 2. Reagents and conditions: (a) allyl bromide, KOtBu, THF/DMSO, 0 °C, 95.2%; (b) Pd(OAc)₂, P(*o*-MePh)₃, Et₃N, aryl bromide, acetonitrile, 60 °C 1 h then 90 °C 24 h; (c) tetra-*n*-butylammonium bromide, anhydrous NaOAc, *trans*-di(μ -acetato)bis[O-(di-*O*-tolylphosphino)benzyl]dipalladium, aryl bromide, DMF, 100 °C 3 days, Ar = 3-pyridyl (b), 5-pyrimidyl (c), 4-isoquinolyl (e); (d) MeOH, 65 °C.



Scheme 3. Reagents and conditions: (a) propargyl bromide, KOtBu, THF/DMSO, 0 °C, 89.5%; (b) (PPh₃)₂PdCl₂, CuI, Et₃N, aryl bromide (or aryl iodide), acetonitrile, 80 °C 3 h, Ar = 4-nitrophenyl (a), 3-pyridyl (b), 5-pyrimidyl (c), 3-quinolyl (d), 4-isoquinolyl (e), 5-indolyl (f); (c) MeOH, 65 °C.

Table 1

In vitro antibacterial activities of alkylides against selected respiratory tract pathogens (MIC, $\mu\text{g/mL}$)

	CLA ^g	AZI ^h	6c	8c	11a	11b	11c	11d	11e
ATCC 49619 ^a	0.032	0.064	0.25	4	32	16	16	16	16
PU-11 ^b	128	128	32	32	32	32	32	16	32
PU-13 ^c	128	128	32	32	32	32	32	16	32
ATCC 29213 ^d	0.5	0.5	8	64	64	64	32	64	128
PU-19 ^e	>256	>256	64	64	128	64	32	128	256
PU-32 ^f	128	64	32	64	32	64	32	64	256

^a *S. pneumoniae* ATCC 49619: erythromycin-susceptible strain.

^b *S. pneumoniae* PU-11: erythromycin-resistant strain bearing both *erm* and *mef* genes.

^c *S. pneumoniae* PU-13: erythromycin-resistant strain bearing both *erm* and *mef* genes.

^d *S. aureus* ATCC 29213: erythromycin-susceptible strain.

^e *S. aureus* PU-19: erythromycin-resistant strain bearing constitutive *erm* (A) gene

^f *S. aureus* PU-32: erythromycin-resistant strain bearing inducible *erm* (A) gene.

^g CLA: clarithromycin.

^h AZI: azithromycin.

against erythromycin-resistant *S. aureus* and *S. pneumoniae* than those of the reference compound, clarithromycin and azithromycin. Among the series of **11**, **11c** (5-pyrimidyl) was the best candidate with 4- to 16-fold greater activities than clarithromycin against the resistant strains, particularly *S. aureus* bearing constitutive *erm* (A) gene. In comparison, the alkylides with fused biaryl rings **11d** (3-quinolyl) and **11e** (4-isoquinolyl) showed inactive against erythromycin-resistant *S. aureus*, despite a slightly higher activities of **11d** than **11c** against erythromycin-resistant *S. pneumoniae* bearing both *erm* and *mef* genes. As for the effect of various sidechain's configuration on the activities, **6c**, **8c**, and **11c** shared similar profile against erythromycin-resistant strains. However, **6c** was 4- to 64-fold more potent than **8c** and **11c** against erythromycin-susceptible strains, particularly 64-fold more potent than **11c** against erythromycin-susceptible *S. pneumoniae*.

In conclusion, we designed and synthesized a series of novel alkylides, possessing three-atom length ether linkers instead of cladinose at 3-OH. Among the alkylides (**6**, **8**, and **11**), **8c** was the promising candidate against erythromycin-resistant strains.

Further SAR studies on alkylides are still in progress, varying the length of the linkers included.

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Supplementary data

Supplementary data associated with this article can be found, in the online version, at [doi:10.1016/j.bmcl.2010.03.038](https://doi.org/10.1016/j.bmcl.2010.03.038).

References and notes

- Kurath, P.; Jones, P. H.; Egan, R. S.; Perun, T. J. *Experientia* **1971**, *27*, 362.
- Watanabe, Y.; Morimoto, S.; Adachi, T.; Kashimura, M.; Asaka, T. *J. Antibiot.* **1993**, *46*, 647.
- Morimoto, S.; Nagate, T.; Sugita, K.; Ono, T.; Numata, K.; Miyachi, J.; Misawa, Y.; Yamada, K.; Omura, S. *J. Antibiot.* **1990**, *43*, 295.
- Katz, L.; Ashley, G. W. *Chem. Rev.* **2005**, *105*, 499.
- Agouridas, C.; Denis, A.; Auger, J. M.; Benedetti, Y.; Bonnefoy, A.; Bretin, F.; Chantot, J. F.; Dussarat, A.; Fromentin, C.; D'Ambrières, S. G.; Lachaud, S.; Laurin, P.; Le Martret, O.; Loyau, V.; Tessot, N. *J. Med. Chem.* **1998**, *41*, 4080.
- Denis, A.; Agouridas, C.; Auger, J. M.; Benedetti, Y.; Bonnefoy, A.; Bretin, F.; Chantot, J. F.; Dussarat, A.; Fromentin, C.; D'Ambrières, S. G.; Lachaud, S.; Laurin, P.; Le Martret, O.; Loyau, V.; Tessot, N.; Pejac, J. M.; Perron, S. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 3075.
- Or, Y. S.; Clark, R. F.; Wang, S.; Chu, D. T. W.; Nilius, A. M.; Flamm, R. K.; Mitten, M.; Ewing, P.; Alder, J.; Ma, Z. *J. Med. Chem.* **2000**, *43*, 1045.
- Kashimura, M.; Asaka, T.; Misawa, Y.; Matsumoto, K.; Morimoto, S. *J. Antibiot.* **2001**, *54*, 664.
- Tanikawa, T.; Asaka, T.; Kashimura, M.; Misawa, Y.; Suzuki, K.; Sato, M.; Kameo, K.; Morimoto, S.; Nishida, A. *J. Med. Chem.* **2001**, *44*, 4027.
- Tanikawa, T.; Asaka, T.; Kashimura, M.; Suzuki, K.; Sugiyama, H.; Sato, M.; Kameo, K.; Morimoto, S.; Nishida, A. *J. Med. Chem.* **2003**, *46*, 2706.
- Asaka, T.; Manaka, A.; Sugiyama, H. *Curr. Top. Med. Chem.* **2003**, *3*, 961.
- Pal, S. *Tetrahedron* **2006**, *62*, 3171.
- Mutak, S. *J. Antibiot.* **2007**, *60*, 85.
- Tang, D.; Gai, Y.; Polemeropoulos, A.; Chen, Z.; Wang, Z.; Or, Y. S. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 5078.
- Elliott, R. L.; Pireh, D.; Griesgraber, G.; Nilius, A. M.; Ewing, P. J.; Bui, M. H.; Raney, P. M.; Flamm, R. K.; Kim, K.; Henry, R. F.; Chu, D. T. W.; Plattner, J. J.; Or, Y. S. *J. Med. Chem.* **1998**, *41*, 1651.
- Denis, A.; Bretin, F.; Fromentin, C.; Bonnet, A.; Piltan, G.; Bonnefoy, A.; Agouridas, C. *Bioorg. Med. Chem. Lett.* **2000**, *10*, 2019.
- Elliott, R. L.; Pireh, D.; Nilius, A. M.; Johnson, P. M.; Flamm, R. K.; Chu, D. T. W.; Plattner, J. J.; Or, Y. S. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 641.
- Misawa, Y.; Asaka, T.; Kashimura, M.; Morimoto, S.; Hatayama, K. E. Patent 0682,038 A1, 1994; *Chem. Abstr.* **122**, 240348.
- Cheng, H. M.; Dirlam, J. P.; Ziegler, C. B.; Lundy, K. M.; Hayashi, S. F.; Kamicker, B. J.; Dutra, J. K.; Daniel, K. L.; Santoro, S. L.; George, D. M.; Bertsche, C. D.; Sakya, S. M.; Suarez-Contreras, M. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 2431.
- Heggelund, A.; Undheim, K. *Tetrahedron Lett.* **2008**, *49*, 5569.
- Clark, R. F.; Ma, Z. K.; Wang, S. Y.; Griesgraber, G.; Tufano, M.; Yong, H.; Li, L. P.; Zhang, X. L.; Nilius, A. M.; Chu, D. T. W.; Or, Y. S. *Bioorg. Med. Chem. Lett.* **2000**, *10*, 815.
- Phan, L. T.; Clark, R. F.; Rupp, M.; Or, Y. S.; Chu, D. T. W.; Ma, Z. *Org. Lett.* **2000**, *2*, 2951.
- Keyes, R. F.; Carter, J. J.; Englund, E. E.; Daly, M. M.; Stone, G. G.; Nilius, A. M.; Ma, Z. K. *J. Med. Chem.* **2003**, *46*, 1795.
- Beebe, X.; Yang, F.; Bui, M. H.; Mitten, M. J.; Ma, Z.; Nilius, A. M.; Djuric, S. W. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 2417.
- Yong, H.; Gu, Y. G.; Clark, R. F.; Marron, T.; Ma, Z.; Soni, N.; Stone, G. G.; Nilius, A. M.; Marsh, K.; Djuric, S. W. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 2653.
- Liang, J. H.; Wang, Y. Y.; Zhu, D. Y.; Dong, L. J.; An, M. M.; Wang, R.; Yao, G. W. *J. Antibiot.* **2009**, *62*, 605.
- Wei, X.; You, Q. D. *Org. Process Res. Dev.* **2006**, *10*, 446.
- Herrmann, W. A.; Brossmer, C.; Reisinger, C. P.; Riermeier, T. H.; Ofele, K.; Beller, M. *Chem. Eur. J.* **1997**, *3*, 1357.
- Furuuchi, T.; Miura, T.; Kurihara, K.; Yoshida, T.; Watanabe, T.; Ajito, K. *Bioorg. Med. Chem.* **2008**, *16*, 4401.
- Sugawara, A.; Sunazuka, T.; Hirose, T.; Nagai, K.; Yamaguchi, Y.; Hanaki, H.; Sharpless, K. B.; Omura, S. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 6340.
- Clinical and Laboratory Standards Institute. *Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically; Approved Standard-Eighth Edition M07-A8*. CLSI, 2009.
- Chen, S. X.; Xu, X. D.; Yu, L. X. *J. Antibiot.* **2001**, *54*, 506.